



THE UNIVERSITY *of* EDINBURGH

Edinburgh Research Explorer

Hemichannel-mediated release of lactate

Citation for published version:

Karagiannis, A, Sylantsev, S, Hadjihambi, A, Hosford, PS, Kasparov, S & Gourine, AV 2015, 'Hemichannel-mediated release of lactate', *Journal of Cerebral Blood Flow and Metabolism*.
<https://doi.org/10.1177/0271678X15611912>

Digital Object Identifier (DOI):

[10.1177/0271678X15611912](https://doi.org/10.1177/0271678X15611912)

Link:

[Link to publication record in Edinburgh Research Explorer](#)

Document Version:

Publisher's PDF, also known as Version of record

Published In:

Journal of Cerebral Blood Flow and Metabolism

Publisher Rights Statement:

This article is distributed under the terms of the Creative Commons Attribution 3.0 License (<http://www.creativecommons.org/licenses/by/3.0/>) which permits any use, reproduction and distribution of the work without further permission provided the original work is attributed as specified on the SAGE and Open Access page (<https://us.sagepub.com/en-us/nam/open-access-at-sage>).

General rights

Copyright for the publications made accessible via the Edinburgh Research Explorer is retained by the author(s) and / or other copyright owners and it is a condition of accessing these publications that users recognise and abide by the legal requirements associated with these rights.

Take down policy

The University of Edinburgh has made every reasonable effort to ensure that Edinburgh Research Explorer content complies with UK legislation. If you believe that the public display of this file breaches copyright please contact openaccess@ed.ac.uk providing details, and we will remove access to the work immediately and investigate your claim.



Hemichannel-mediated release of lactate

Anastassios Karagiannis¹, Sergiy Sylantjev², Anna Hadjihambi¹,
Patrick S Hosford¹, Sergey Kasparov³
and Alexander V Gourine¹

Abstract

In the central nervous system lactate contributes to the extracellular pool of readily available energy substrates and may also function as a signaling molecule which mediates communication between glial cells and neurons. Monocarboxylate transporters are believed to provide the main pathway for lactate transport across the membranes. Here we tested the hypothesis that lactate could also be released via opening of pannexin and/or functional connexin hemichannels. In acute slices prepared from the brainstem, hippocampus, hypothalamus and cortex of adult rats, enzymatic amperometric biosensors detected significant tonic lactate release inhibited by compounds, which block pannexin/connexin hemichannels and facilitated by lowering extracellular $[Ca^{2+}]$ or increased PCO_2 . Enhanced lactate release triggered by hypoxia was reduced by ~50% by either connexin or monocarboxylate transporter blockers. Stimulation of Schaffer collateral fibers triggered lactate release in CA1 area of the hippocampus, which was facilitated in conditions of low extracellular $[Ca^{2+}]$, markedly reduced by blockade of connexin hemichannels and abolished by lactate dehydrogenase inhibitor oxamate. These results indicate that lactate transport across the membranes may occur via mechanisms other than monocarboxylate transporters. In the central nervous system, hemichannels may function as a conduit of lactate release, and this mechanism is recruited during hypoxia and periods of enhanced neuronal activity.

Keywords

Astrocytes, connexin, lactate, metabolism, monocarboxylate transporters, pannexin

Received 31 May 2015; Revised 14 September 2015; Accepted 21 September 2015

Introduction

Neurons require constant and sufficient supply of energy substrates to meet their metabolic needs. Lactate is one of the key elements of brain energy metabolism and contributes to the extracellular pool of readily available energy substrates. The so-called ‘astrocyte-to-neuron lactate shuttle’ hypothesis suggests that neuronal activity is fuelled by lactate provided by neighbouring astrocytes.^{1,2} Although there is evidence which argues against the importance (and existence) of astrocyte-to-neuron lactate shuttle (for a comprehensive review see Dienel³), the role of lactate and its effective transport in supporting metabolic needs of the brain may become highly significant in certain conditions. For example, physical activity and exercise are associated with progressive increase in brain lactate uptake (from the blood) and oxidation as work load and plasma lactate levels rise.^{4,5} In addition to its metabolic role, lactate may also function as a signaling

molecule which in the brain mediates communication between astrocytes and neurons.^{6–9}

In order to fulfil metabolic (at rest or during exercise) and/or signaling roles, lactate requires an effective mechanism of transport across the membrane(s) – the process which is currently attributed to the operation of monocarboxylate transporters (MCTs).¹⁰ This, however, does not exclude the involvement of other

¹Department of Neuroscience, Physiology and Pharmacology, Centre for Cardiovascular and Metabolic Neuroscience, University College London (UCL), London, UK

²Centre for Clinical Brain Sciences, University of Edinburgh, Edinburgh, UK

³Department of Physiology and Pharmacology, University of Bristol, Bristol, UK

Corresponding author:

Alexander V Gourine, Department of Neuroscience, Physiology & Pharmacology, University College London, WC1E 6BT, UK.
Email: a.gourine@ucl.ac.uk

potential mechanisms.¹¹ Gap junction hemichannels have been reported to be permeable to various molecules including adenosine 5'-triphosphate (ATP), glutamate and glucose,^{12–18} and in this study we tested the hypothesis that lactate could be released into the extracellular space via opening of pannexin and/or functional (i.e. unopposed) gap junction connexin hemichannels.

Materials and methods

All the experiments were performed in accordance with the European Commission Directive 2010/63/EU (European Convention for the Protection of Vertebrate Animals used for Experimental and Other Scientific Purposes) and the UK Home Office (Scientific Procedures) Act (1986) with project approval from the UCL Institutional Animal Care and Use Committee.

In vitro slice preparations

Young adult Sprague-Dawley rats (~120 g) were terminally anesthetized with halothane inhalation overdose, the brains were removed and placed in ice cold artificial cerebrospinal fluid (aCSF) containing 124 mM NaCl, 26 mM NaHCO₃, 3 mM KCl, 2 mM CaCl₂, 1.25 mM NaH₂PO₄, 1 mM MgSO₄, 10 mM Glucose

saturated with 95% O₂/5% CO₂ (pH 7.4) with an addition of 9 mM Mg²⁺. Horizontal brainstem slices containing the intact ventral surface (thickness ~400 μm) or coronal brainstem, hypothalamic or cortical slices (thickness 300 μm) were cut using a vibratome and then incubated at room temperature for 1 h in a standard aCSF saturated with 95% O₂/5% CO₂.

Biosensor recordings

Release of lactate was recorded in real-time using amperometric enzymatic lactate biosensors (Sarissa Biomedical) placed in a direct contact with the surface of the slice. The operation of the lactate biosensor is based on enzymatic activity of lactate oxidase which, in the presence of oxygen, converts lactate to pyruvate and H₂O₂. The latter is detected electrochemically. Figure 1 illustrates linearity of lactate detection by the biosensors in concentrations from 10 μM to 500 μM and insensitivity of the lactate biosensor detection system to pharmacological agents used in the current study. A dual recording configuration of a null sensor (lacking lactate oxidase) and lactate biosensor was used (Figure 2(a)), as described previously.^{19–22} The null sensor served as a control to determine whether any non-specific electroactive interferents were released

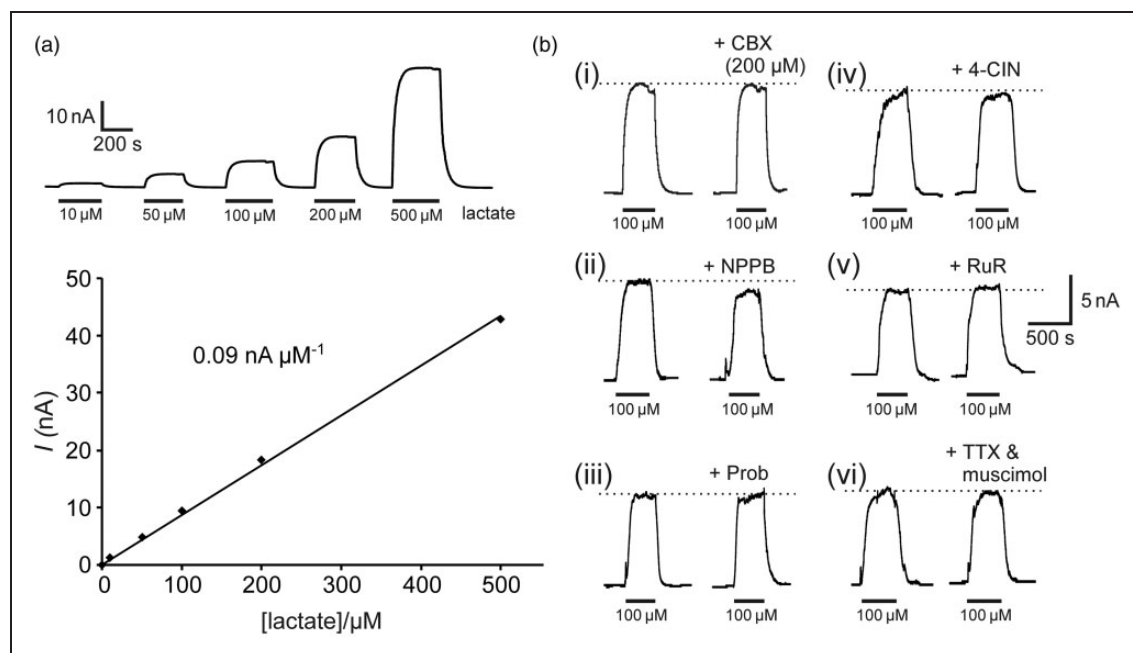
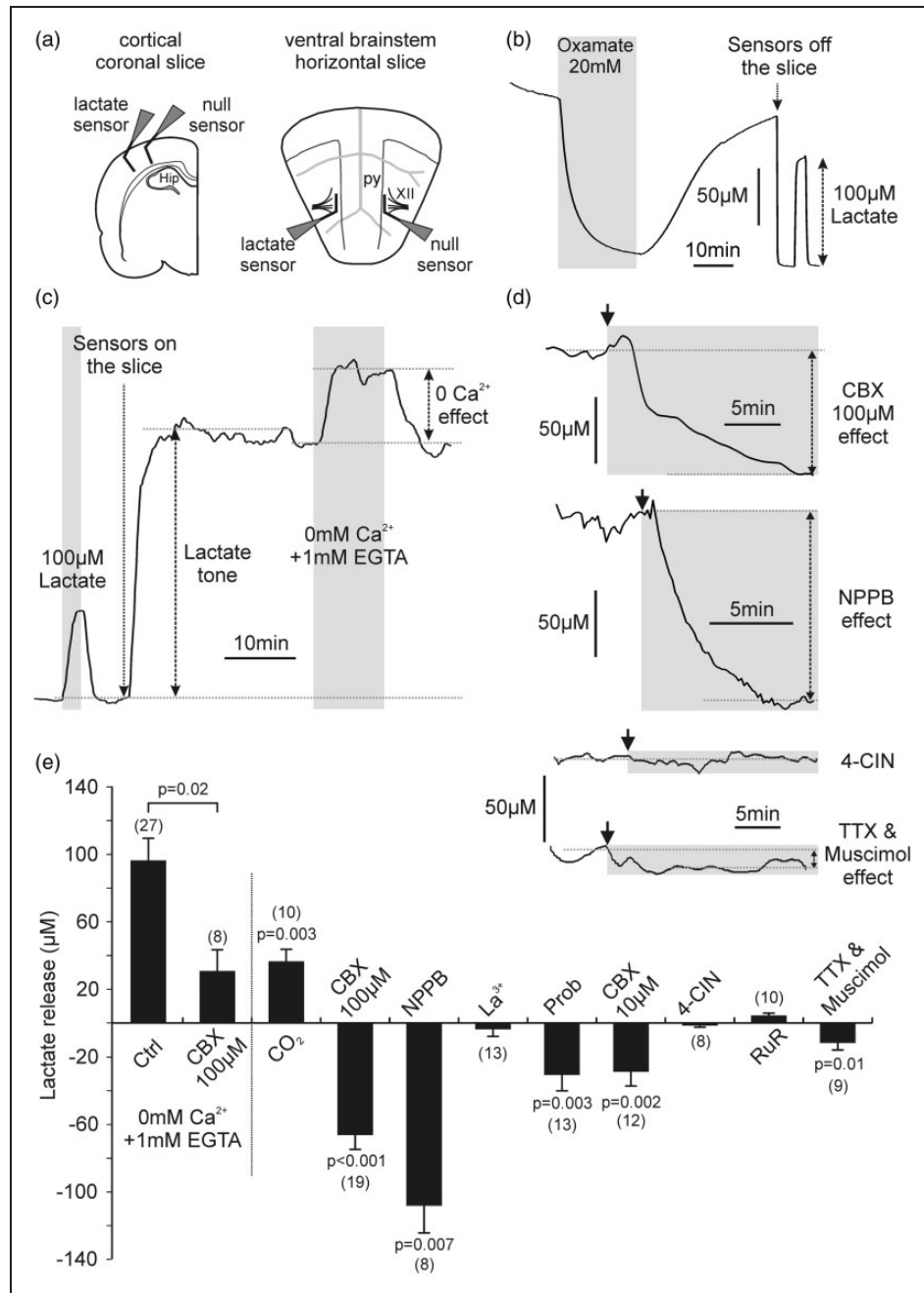


Figure 1. Performance of the lactate biosensors. (a) Current (I) calibration curve and sample responses of the lactate biosensor demonstrating linearity of analyte detection in concentrations from 10 μM to 500 μM; (b) Insensitivity of the lactate biosensor detection system to pharmacological agents used in this study. Traces illustrate sample biosensor current responses to lactate (100 μM) when calibrated in control artificial cerebrospinal fluid (aCSF), and then in the presence of (i) carbenoxolone (CBX), (ii) 5-nitro-2-(3-phenylpropylamino)-benzoic acid (NPPB, 200 μM), (iii) probenecid (Prob, 150 μM), (iv) α-cyano-4-hydroxycinnamate (4-CIN, 250 μM), (v) Ruthenium red (RuR, 20 μM), and (vi) tetrodotoxin (TTX, 1 μM) + muscimol (100 μM).



and confound the measurements. Null sensor currents were subtracted from the lactate biosensor currents. Biosensors were calibrated with known amount of lactate (10 or 100 μM) directly in the slice chamber (i.e. in the identical temperature, aCSF composition/osmolarity conditions) immediately before and after the recordings (Figure 2(b) and (c)). To convert changes in sensor current to changes in lactate concentration, the means of the initial and final calibrations were used.

Recordings were made from the slice placed on an elevated grid in a flow chamber at 35°C. Isohydic hypercapnia was mimicked by replacing normal aCSF with a solution containing 70 mM NaCl, 80 mM NaHCO_3 , 3 mM KCl, 2 mM CaCl_2 , 1.25 mM NaH_2PO_4 , 1 mM MgSO_4 , 10 mM Glucose saturated with 12% CO_2 (PCO_2 70–80 mmHg, pH 7.4). Hypoxic conditions were induced for 2–4 min by replacement of oxygen in the medium with nitrogen (perfusion of the chamber with aCSF saturated with 95% N_2 /5% CO_2). The effect of hypoxia was determined by measuring the peak lactate release upon re-oxygenation (Figure 3(a)) as previously

described.²² This represents an accurate estimation of peak hypoxia-induced lactate release since similar release is triggered by application of a mitochondrial complex III inhibitor myxothiazol (2.5 μM) (Figure 3(b) and (c)).

Electrophysiology

Coronal hippocampal slices (250 μm) were cut from the brains of 25–30-day-old Sprague-Dawley rats and incubated for 1 h in normal aCSF saturated with 95% O_2 /5% CO_2 (pH 7.4). Recordings were made in aCSF containing 2 mM CaCl_2 and 2 mM MgCl_2 at 33–35°C. AMPA-receptor (AMPA)-mediated currents were isolated by adding CGP-55845 (1 μM), D-APV (50 μM), strychnine (1 μM), picrotoxin (100 μM) and S-MCPG (200 μM). The pipette solution contained 117.5 mM Cs-gluconate, 17.5 mM CsCl, 10 mM KOH-HEPES, 10 mM BAPTA, 8 mM NaCl, 5 mM QX-314, 2 mM Mg-ATP, 0.3 mM GTP (pH 7.2, 295 mOsm, resistance 7–9 M Ω). Damaged cells from the slice surface were removed by applying flow of pressurized

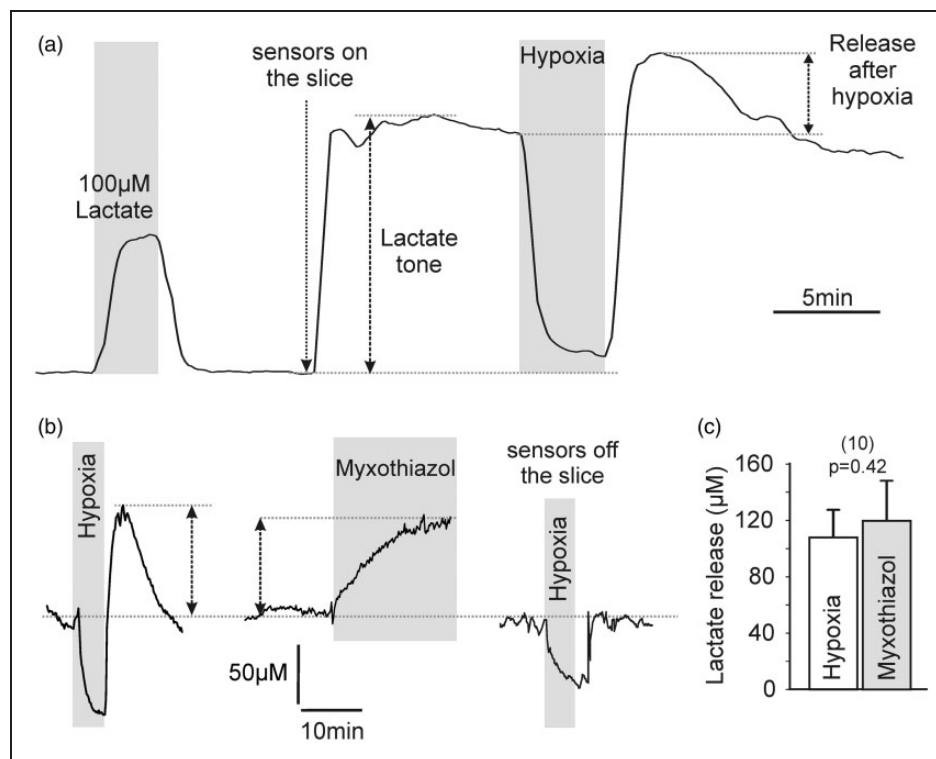


Figure 3. Facilitated release of lactate in response to hypoxia. (a) Representative example of changes in lactate biosensor current during calibration, after biosensor placement on the surface of the brainstem slice, and in response to a hypoxic challenge (perfusion with aCSF saturated with 95% N_2 /5% CO_2). Peak hypoxia-induced lactate release is measured upon re-oxygenation. (b) Representative examples of changes in lactate biosensor current recorded during and after hypoxia when the sensor is placed on the surface of the brain slice (left) or in the bath away from the slice (right) and on the surface of the slice during application of myxothiazol (middle). Decrease in O_2 availability reduces biosensor current followed by a positive signal upon re-oxygenation (hypoxia-induced lactate release), which is only observed when the sensor is in a direct contact with the brain tissue. (c) Peak lactate efflux of a similar magnitude is triggered by hypoxia and myxothiazol.

perfusion solution delivered via a glass pipette (diameter 7–10 μm). Whole-cell voltage-clamp recordings were taken from CA1 pyramidal neurons. Signals were digitized at 10 kHz. Stimulation was applied using a bipolar tungsten electrode placed in the Schaffer collaterals area. Electrical stimuli were delivered every 15 s for 10 min in each experimental episode (application of drugs, change of extracellular $[\text{Ca}^{2+}]$); 10 min intervals without stimulation were given between experimental episodes. Miniature ($\sim 20 \mu\text{m}$ tip diameter) carbon fiber lactate biosensors were placed in the vicinity (within 15–20 μm) of the recorded neurons. Readings of the biosensor currents were taken every 0.5 s, and were synchronized with the recordings of the whole-cell AMPAR-mediated EPSCs.

Drugs

Carbenoxolone (CBX) in 10 μM concentration and probenecid (150 μM) were used to inhibit pannexin hemichannel activity. CBX (100–200 μM), 5-nitro-2-(3-phenylpropylamino)-benzoic acid (NPPB) (200 μM) and La^{3+} (100 μM) were used to interfere with connexin hemichannels. MCTs were inhibited with 4-CIN (250–500 μM). Ruthenium red (20 μM) was used to block CALHM1 channels. Tetrodotoxin (1 μM) and muscimol (100 μM) were applied to inhibit neuronal activity. EGTA (1 mM) was used to chelate Ca^{2+} . All drugs were from Sigma (Poole, UK).

Data analysis

Lactate biosensor measurements were processed using 1401 interface and analyzed using *Spike 2* software (Cambridge Electronic Design). Data are reported as mean \pm S.E.M. Datasets were compared using paired samples Wilcoxon Signed Ranks Test or Student's paired or unpaired *t* test, as appropriate. Differences between groups with *p* values of less than 0.05 were considered significant.

Results

Enzymatic amperometric lactate biosensors were used for real-time recordings of tonic-, hypoxia- and enhanced neuronal activity-stimulated release of lactate in acute slices of rat brainstem, hypothalamus, hippocampus and cortex.

Hemichannel-mediated tonic release of lactate

Tonic lactate efflux (lactate tone; Figure 2(c)) was detected when lactate biosensors were placed in a direct contact with the uncut ventral surface of the brainstem horizontal slices ($278 \pm 12 \mu\text{M}$; $n = 100$,

$p < 0.001$), coronal hypothalamic slices ($349 \pm 46 \mu\text{M}$; $n = 8$, $p = 0.007$), or coronal cortical slices ($229 \pm 15 \mu\text{M}$; $n = 10$, $p = 0.003$). Lactate tone detected on the slice surface was dramatically reduced by application of lactate dehydrogenase inhibitor oxamate (20 mM) (reduction in tone by $-76 \pm 7\%$, $n = 19$, $p < 0.001$, Figure 2(b)), confirming the analyte selectivity of the biosensor detection system.

Opening probability of hemichannels increases when extracellular $[\text{Ca}^{2+}]$ decreases.²³ Lowering extracellular $[\text{Ca}^{2+}]$ (from 2 mM to 0 mM with an addition of 1 mM EGTA to chelate residual Ca^{2+}) facilitated release of lactate by the brainstem (by $94 \pm 15 \mu\text{M}$; $n = 22$, $p = 0.001$) and cortical (by $109 \pm 15 \mu\text{M}$; $n = 5$, $p = 0.03$) slices (Figure 2(c) and (e)). Opening probability of certain connexin hemichannels (e.g. connexin26) also increases in response to CO_2 .^{15,24,25} Isohydic hypercapnia (application of aCSF containing 80 mM HCO_3^- , PCO_2 70–80 mmHg, pH 7.4) facilitated lactate release from the ventral surface of the brainstem (by $37 \pm 7 \mu\text{M}$; $n = 10$, $p = 0.003$, Figure 2(e)). These data suggested that hemichannels may act as a conduit for lactate release.

To further investigate the role of pannexin/connexin hemichannels in mediating tonic release of lactate, we next applied compounds known to inhibit hemichannel activity. CBX and NPPB which block both pannexin and connexin hemichannels significantly reduced lactate tone (Figure 2(d) and (e)). CBX (100 μM) decreased lactate tone by $65 \pm 12 \mu\text{M}$ ($n = 14$, $p < 0.001$) in the brainstem slices and by $71 \pm 7 \mu\text{M}$ ($n = 5$, $p = 0.03$) in the cortical slices. NPPB (200 μM) had a similar effect in the brainstem slices (decrease in lactate tone by $108 \pm 17 \mu\text{M}$ (35%); $n = 8$, $p = 0.007$). The amount of lactate released in response to lowering extracellular $[\text{Ca}^{2+}]$ was also markedly reduced in the presence of CBX ($-64 \pm 22\%$, $n = 8$, $p = 0.02$; Figure 2(e)).

Lactate tone was not affected by La^{3+} (100 μM , Figure 2(e)) which is believed to inhibit connexin, but not pannexin hemichannels,^{26,27} suggesting that the latter might be involved. Indeed, inhibition of pannexins by application of CBX at a low concentration (10 μM) or probenecid (150 μM) decreased lactate tone (CBX 10 μM : $-29 \pm 9 \mu\text{M}$, $n = 12$, $p = 0.002$; probenecid: $-31 \pm 9 \mu\text{M}$, $n = 13$, $p = 0.003$) (Figure 2(e)). Interestingly, inhibitor of MCTs 4-CIN (250–500 μM) had no effect on tonic release of lactate by either brainstem or cortical slices (Figure 2(d) and (e)). Inhibition of neuronal activity by co-application of tetrodotoxin (TTX, 1 μM) and muscimol (100 μM) was associated with a small reduction in lactate tone ($-12 \pm 4 \mu\text{M}$; $n = 9$, $p = 0.01$, Figure 2(d) and (e)). Application of ruthenium red (20 μM) to block CALHM1 channels (which are also sensitive to low extracellular $[\text{Ca}^{2+}]$)²⁸

did not reduce the lactate tone (increase by $4.4 \pm 1.6 \mu\text{M}$, $n = 10$, Figure 2(e)).

Control tests were performed to determine whether the pharmacological agents used in this study have an effect on lactate biosensor detection system (Figure 1(b)). Only NPPB (200 μM) was found to reduce biosensor current by $\sim 10\%$, which is significantly less compared to the effect of the drug on currents generated by the biosensors when positioned on the brain slice (reduction in lactate tone by 35% and reduction in hypoxia-induced lactate release by $> 50\%$, see below).

Connexin hemichannels and MCTs mediate facilitated release of lactate during hypoxia

Tissue hypoxia decreases mitochondrial utilization of pyruvate leading to an increase in lactate production and release. Since biosensors require oxygen to operate (*lactate oxidase* entrapped in the biosensor polymer layer converts lactate to pyruvate and H_2O_2 which is detected electrochemically), decreases in O_2 concentration (saturation of the media with 95% $\text{N}_2/5\% \text{CO}_2$) reduce biosensor currents (Figure 3(a) and (b)). Therefore, lactate release induced by hypoxia can only be determined upon re-oxygenation²² (increase in lactate release in the brainstem slices from $219 \pm 16 \mu\text{M}$ to $284 \pm 18 \mu\text{M}$; $n = 61$; $p < 0.001$; in the cortical slices from $182 \pm 10 \mu\text{M}$ to $302 \pm 26 \mu\text{M}$; $n = 5$; $p = 0.03$). These values represent an accurate estimation of peak hypoxia-induced lactate release since lactate efflux of a similar magnitude is triggered by bath application of the mitochondrial complex III inhibitor myxothiazol (Figure 3(b) and (c)). Hypoxia-induced lactate release was not affected by CBX applied at a low concentration (10 μM) (Figure 4(b)) or probenecid, suggesting that pannexins are not involved. However, lactate release triggered by hypoxia was significantly (by $\sim 50\%$) reduced in the presence of either MCTs inhibitor 4-CIN (250 μM , $p = 0.03$) or any of the tested connexin hemichannel blockers (CBX 100 μM , $p = 0.006$; NPPB, $p = 0.007$; or La^{3+} , $p = 0.001$) (Figure 4(a) and (b)). Blockade of the neuronal activity with TTX and muscimol resulted in a slightly higher peak lactate release in response to hypoxic challenge (Figure 4(a) and (b)).

Connexin hemichannels contribute to the release of lactate triggered by enhanced neuronal activity

We next examined hemichannel involvement in the release of lactate driven by enhanced neuronal activity.²⁹ Miniature ($\sim 20 \mu\text{m}$ in diameter) carbon fiber lactate biosensors placed in the vicinity of the patch-clamped hippocampal CA1 neurons (Figure 5(a)) recorded significant (peak release $35 \pm 3 \mu\text{M}$, $n = 11$) lactate release in response to stimulation of Schaffer

collateral fibers (Figure 5(b) and (d)). Lactate release in CA1 in response to stimulation of Schaffer collaterals was increased in low extracellular $[\text{Ca}^{2+}]$ (0.2 mM) conditions (peak release $48 \pm 4 \mu\text{M}$, $n = 5$, $p < 0.05$; Figure 5(b) and (d)), abolished in the presence of oxamate applied 20 min prior ($n = 5$, $p < 0.001$; Figure 5(b) and (d)), and markedly reduced by CBX (peak release $12 \pm 2 \mu\text{M}$, $n = 6$; $p < 0.001$; Figure 5(d)). Simultaneous recordings of AMPA-receptor-mediated EPSCs showed that decreased release of lactate in the presence of CBX is associated with small decreases in the amplitude of the evoked EPSCs (Figure 5(c) and (e)). CBX applied at a low concentration (10 μM) had no effect on the evoked EPSCs and activity-dependent lactate release (Figure 5(c) to (e)).

Discussion

Lactate is continuously produced by the brain parenchyma even in conditions of ample oxygen supply.³⁰ Glycolysis in the presence of oxygen (aerobic glycolysis) is an interesting feature of brain metabolism, and in resting conditions there is a net efflux of lactate from the CNS, i.e. lactate concentration in the venous blood is higher than in the arterial blood supplying the brain.^{31,32} For decades it has been thought that MCTs are the only pathway of lactate transport from the cells which produce it. This dogma was challenged by the results of the recent study suggesting that lactate release may occur via the mechanisms involving certain membrane channels.¹¹ The data obtained in the present study suggest that these alternative mechanisms include pannexin and functional connexin hemichannels. We report that hemichannels are capable to function as a conduit of lactate release and this mechanism is recruited during hypoxia and periods of enhanced neuronal activity.

We show that tonic efflux of lactate can indeed be detected in hyperoxygenated environment ($\text{PO}_2 > 500 \text{ mmHg}$ in standard conditions used for the experiments in brain slices superfused with aCSF saturated with 95% $\text{O}_2/5\% \text{CO}_2$) in reduced preparations, such as acute brainstem, hypothalamic or cortical slices. Lactate tone detected on the intact ventral brainstem surface was similar to that recorded by the biosensors placed in a direct contact with coronal brain sections, indicating that damaged cells are not the primary source of this release. Lactate tone increased in conditions of low extracellular $[\text{Ca}^{2+}]$ or high PCO_2 , which favor hemichannel opening,^{15,24,25,33} and decreased in the presence of pharmacological agents known to inhibit pannexin and connexin hemichannels. Efflux of lactate facilitated by hypoxia was significantly reduced by either connexin or MCT blockers. Lactate release in CA1 triggered in response to stimulation of

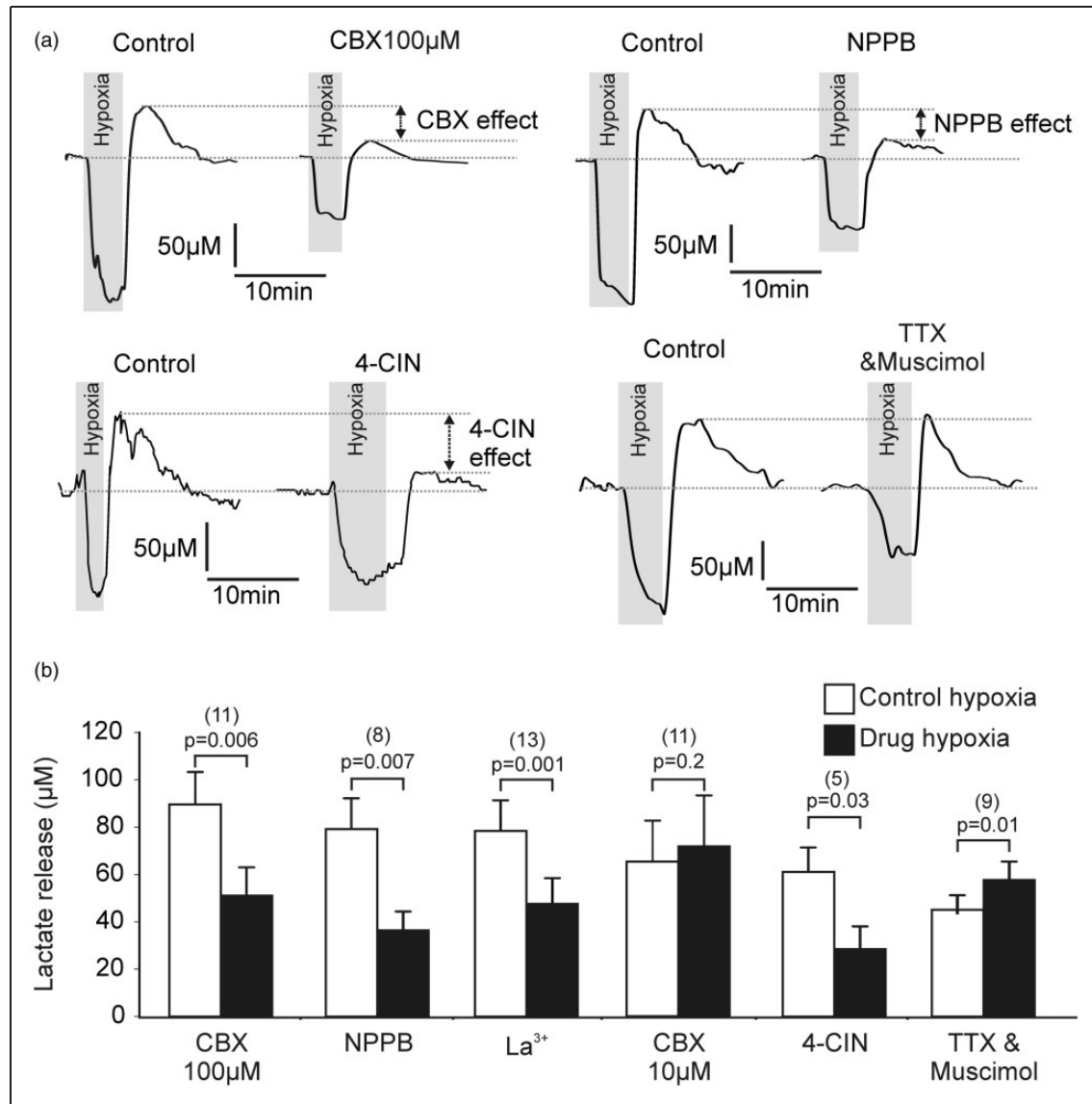


Figure 4. Connexin hemichannels and monocarboxylate transporters (MCTs) mediate facilitated release of lactate during hypoxia. (a) Representative recordings of changes in lactate biosensor current showing hypoxia-induced lactate release by the brainstem slices in the absence and presence of CBX (100 μM), NPPB (200 μM), 4-CIN (250 μM) or co-application of TTX (1 μM) and muscimol (100 μM); (b) Summary data illustrating peak hypoxia-induced lactate release in the absence and presence of CBX, NPPB, La³⁺, 4-CIN, TTX+muscimol.

Schaffer collaterals was enhanced in low extracellular [Ca²⁺] and reduced by CBX. This is interesting because lowering extracellular [Ca²⁺] should reduce the synaptic strength and affect neuronal responses in the projection area. This observation suggests that release of lactate might be uncoupled from synaptic activity responding to ionic movements associated with axonal and dendritic membrane potentials propagation. For example, there is recent evidence showing that an increase in extracellular concentration of K⁺ (within the physiological range) is sufficient to trigger release of lactate by astrocytes.¹¹ Through hemichannels or other

unidentified channels,¹¹ the lactate anion (Lac⁻) can exit cells without the involvement of the proton and facilitate Na⁺ entry (through the activity of sodium-coupled MCTs), which is required by the Na⁺/K⁺ATPase for uptake of neuronally released K⁺.

It needs to be acknowledged that none of the pharmacological compounds available today are specific hemichannel blockers. Nevertheless, they all have different off target effects strongly suggesting that hemichannels are responsible for this lactate release. Facilitated lactate release in response to lowering extracellular [Ca²⁺] or increases in tissue PCO₂ further supports this conclusion.

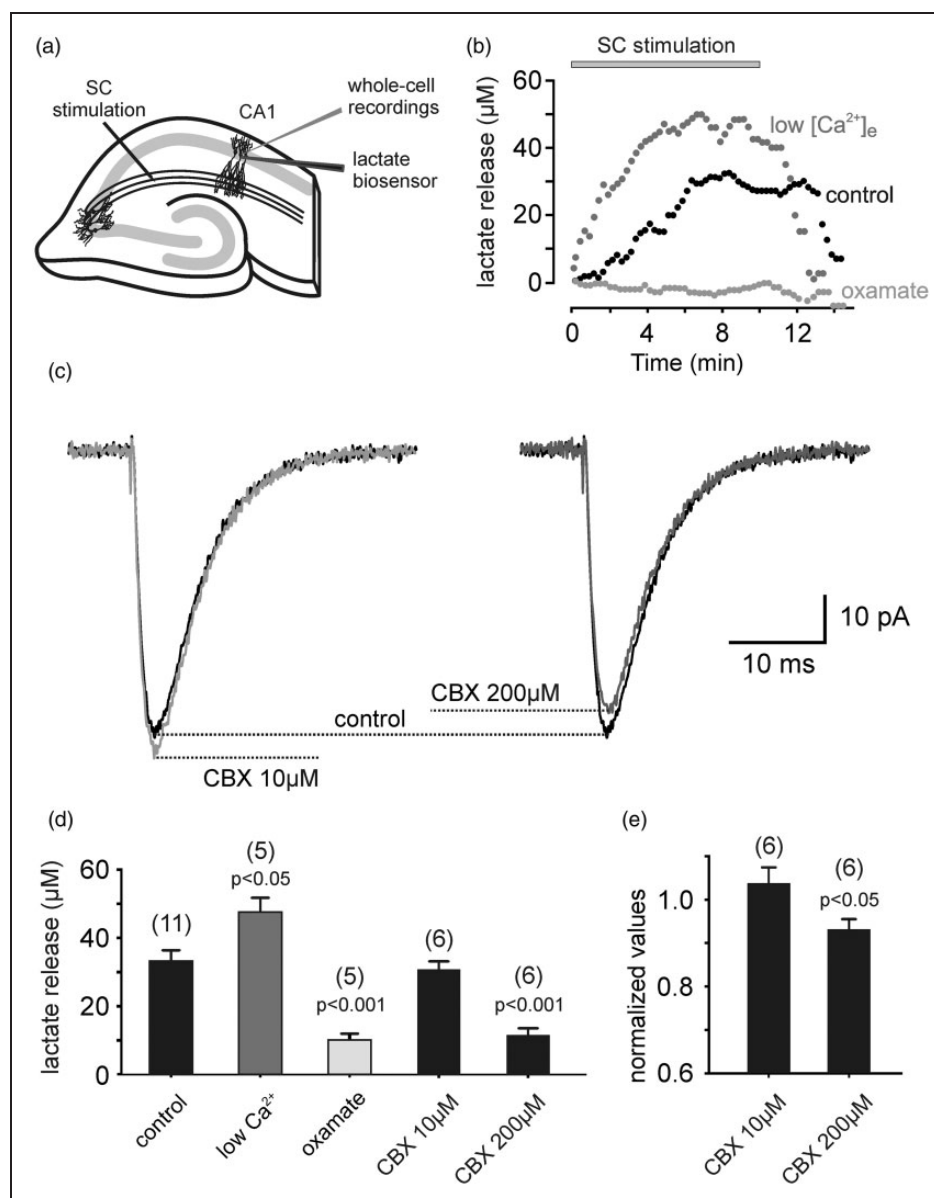


Figure 5. Connexin hemichannels contribute to the increases in lactate release triggered by enhanced neuronal activity. (a) Schematic drawing of the experimental design. Miniature (20 μm) carbon fiber lactate biosensors were placed in the vicinity of the patch-clamped hippocampal CA1 neurons to record lactate release in response to electrical stimulation of Schaffer collateral (SC) fibers; (b) Representative recordings of changes in biosensor current illustrating lactate release in CA1 in response to stimulation of Schaffer collaterals in control and low (0.2 mM) extracellular $[Ca^{2+}]_e$ conditions and in the presence of oxamate (20 mM). Schaffer collaterals were stimulated for 10 min; (c) Representative AMPAR EPSCs recorded in the same experiment in the absence and presence of CBX (each trace is an average of 10); (d) Summary data illustrating peak lactate release in CA1 area in response to stimulation of Schaffer collaterals in control and low extracellular $[Ca^{2+}]_e$ conditions, in the presence of oxamate or CBX; (e) Summary data illustrating amplitudes of AMPAR-mediated EPSCs normalized to the control values.

Our pharmacological survey also suggests that other membrane channels sensitive to changes in extracellular $[Ca^{2+}]_e$ (e.g. CALHM1, effectively blocked by Ruthenium red) are unlikely to be involved.

These results provide strong evidence that MCTs are not the sole conduit of lactate release in the CNS. It is rather surprising that the evidence of membrane

channel-mediated lactate release was only recently reported,¹¹ considering that molecules as large as ATP and glucose are known to be transported via hemichannel opening.^{12–18,25} All cells of the brain parenchyma (neurons, astrocytes, oligodendrocytes and microglia) express pannexins and connexins and identification of the exact source(s) of lactate release detected by the

biosensors on the surface of the brain slice *in vitro* is beyond the scope of the present study. Nevertheless, the existing evidence points to astroglia as an important source of lactate release. Recent studies involving measurements of cytosolic lactate concentration in astrocytes using genetically encoded fluorescence sensors reported values of ~ 1.4 mM and concluded that at resting conditions astrocytes maintain a steady-state reservoir of lactate allowing its rapid mobilization.¹¹ Pharmacological compounds which are expected to significantly reduce neuronal activity (TTX and muscimol) had only a small effect on tonic lactate release, while astroglial activity appears to correlate well with lactate production and release by these cells.^{2,6,11}

In summary, we show that pannexin and/or connexin hemichannels are capable of functioning as a conduit of lactate transport across the membrane and this mechanism is recruited in conditions of low PO_2 , high PCO_2 or during periods of enhanced neuronal activity. The identified mechanism may play a role in providing rapid and effective local supply of lactate to meet increased metabolic demands of active neurons. Indeed, extracellular $[Ca^{2+}]$ can decrease to ~ 0.7 mM during periods of intense neuronal activity.^{34–37} This would increase opening probability of hemichannels expressed by neighbouring astrocytes leading to highly localized neuronal activity-driven release of lactate “on-site”. Neuronal activity-dependent opening of connexin hemichannels triggered by decreases in extracellular $[Ca^{2+}]$ (and leading to the release of ATP) has been demonstrated previously.³⁸ At rest, hemichannel-mediated mechanisms may contribute to the convective flux of lactate across the astrocytic syncytium and its clearance from the brain via the circulation³ and/or the glymphatic system.³⁹ During physical activity, when peripheral lactate is fuelling our brains, hemichannels are hypothesized to play an important role in the effective uptake of this metabolic substrate from the arterial blood.

Funding

This study was supported by The Wellcome Trust. A.V.G is a Wellcome Trust Senior Research Fellow (ref. 095064). S.S. research is funded by Moray Endowment Grant and Wellcome Trust ISSF-2 Grant 217SSV.

Declaration of conflicting interests

The author(s) declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

Authors' contributions

A.V.G. and S.S. conceived and designed the experiments. A.K., S.S., A.H. and P.S.H. collected, assembled and analyzed the data. A.V.G., S.S. and S.K. interpreted the data. A.V.G. and S.K. wrote the article. All authors revised the article critically for important intellectual content.

References

- Pellerin L and Magistretti PJ. Sweet sixteen for ANLS. *J Cereb Blood Flow Metab* 2012; 32: 1152–1166.
- Pellerin L and Magistretti PJ. Glutamate uptake into astrocytes stimulates aerobic glycolysis: a mechanism coupling neuronal activity to glucose utilization. *Proc Natl Acad Sci U S A* 1994; 91: 10625–10629.
- Dienel GA. Brain lactate metabolism: the discoveries and the controversies. *J Cereb Blood Flow Metab* 2012; 32: 1107–1138.
- Ide K, Horn A and Secher N. Cerebral metabolic response to submaximal exercise. *J Appl Physiol* 1999; 87: 1604–1608.
- Ide K, Schmalbruch IK, Quistorff B, et al. Lactate, glucose and O_2 uptake in human brain during recovery from maximal exercise. *J Physiol* 2000; 522: 159–164.
- Tang F, Lane S, Korsak A, et al. Lactate-mediated glia-neuronal signalling in the mammalian brain. *Nat Commun* 2014; 5: 3284.
- Yang J, Ruchti E, Petit J-MJ-M, et al. Lactate promotes plasticity gene expression by potentiating NMDA signaling in neurons. *Proc Natl Acad Sci* 2014; 111: 12228–12233.
- Mosienko V, Teschemacher AG and Kasparov S. Is L-lactate a novel signaling molecule in the brain? *J Cereb Blood Flow Metab* 2015; 35: 1069–1075.
- Teschemacher AG, Gourine AV and Kasparov S. A role for astrocytes in sensing the brain microenvironment and neuro-metabolic integration. *Neurochem Res* 2015; 1–8. doi:10.1007/s11064-015-1562-9.
- Halestrap AP and Wilson MC. The monocarboxylate transporter family-role and regulation. *IUBMB Life* 2012; 64: 109–119.
- Sotelo-Hitschfeld T, Niemeyer MI, Machler P, et al. Channel-mediated lactate release by K^+ -stimulated astrocytes. *J Neurosci* 2015; 35: 4168–4178.
- Rouach N, Koulakoff A, Abudara V, et al. Astroglial metabolic networks sustain hippocampal synaptic transmission. *Science* 2008; 322: 1551–1555.
- Kang J, Kang N, Lovatt D, et al. Connexin 43 hemichannels are permeable to ATP. *J Neurosci* 2008; 28: 4702–4711.
- Sáez JC and Leybaert L. Hunting for connexin hemichannels. *FEBS Letters* 2014; 588: 1205–1211.
- Huckstepp RTR, id Bihi R, Eason R, et al. Connexin hemichannel-mediated CO_2 -dependent release of ATP in the medulla oblongata contributes to central respiratory chemosensitivity. *J Physiol* 2010; 588: 3901–3920.
- Ye Z, Wyeth MS, Baltan-tekkok S, et al. Functional hemichannels in astrocytes: a novel mechanism of glutamate release. *J Neurosci* 2003; 23: 3588–3596.
- Harris AL. Connexin channel permeability to cytoplasmic molecules. *Prog Biophys Mol Biol* 2007; 94: 120–143.
- Tabernero A, Giaume C and Medina JM. Endothelin-1 regulates glucose utilization in cultured astrocytes by controlling intercellular communication through gap junctions. *Glia* 1996; 16: 187–195.
- Gourine AV, Llaudet E, Thomas T, et al. Adenosine release in nucleus tractus solitarius does not appear to

- mediate hypoxia-induced respiratory depression in rats. *J Physiol* 2002; 544: 161–170.
20. Gourine AV, Dale N, Korsak A, et al. Release of ATP and glutamate in the nucleus tractus solitarius mediate pulmonary stretch receptor (Breuer–Hering) reflex pathway. *J Physiol* 2008; 16: 3963–3978.
 21. Turovsky E, Karagiannis A, Abdala AP, et al. Impaired CO₂ sensitivity of astrocytes in a mouse model of Rett syndrome. *J Physiol* 2015; 593: 3159–3168.
 22. Wells JA, Christie IN, Hosford PS, et al. A critical role for purinergic signalling in the mechanisms underlying generation of BOLD fMRI responses. *J Neurosci* 2015; 35: 5284–5292.
 23. Müller DJ, Hand GM, Engel A, et al. Conformational changes in surface structures of isolated connexin 26 gap junctions. *EMBO J* 2002; 21: 3598–3607.
 24. Huckstepp RTR, Eason R, Sachdev A, et al. CO₂-dependent opening of connexin 26 and related β connexins. *J Physiol* 2010; 588: 3921–3931.
 25. Meigh L, Greenhalgh SA, Rodgers TL, et al. CO₂ directly modulates connexin 26 by formation of carbamate bridges between subunits. *Elife* 2013; 2: e01213.
 26. Orellana JA, Froger N, Ezan P, et al. ATP and glutamate released via astroglial connexin43 hemichannels mediate neuronal death through activation of pannexin 1 hemichannels. *J Neurochem* 2011; 118: 826–840.
 27. Orellana JA, Hernández DE, Ezan P, et al. Hypoxia in high glucose followed by reoxygenation in normal glucose reduces the viability of cortical astrocytes through increased permeability of connexin 43 hemichannels. *Glia* 2010; 58: 329–343.
 28. Taruno A, Vingtdoux V, Ohmoto M, et al. CALHM1 ion channel mediates purinergic neurotransmission of sweet, bitter and umami tastes. *Nature* 2013; 495: 223–226.
 29. Hu Y and Wilson GS. A temporary local energy pool coupled to neuronal activity: fluctuations of extracellular lactate levels in rat brain monitored with rapid-response enzyme-based sensor. *J Neurochem* 1997; 69: 1484–1490.
 30. Abi-Saab WM, Maggs DG, Jones T, et al. Striking differences in glucose and lactate levels between brain extracellular fluid and plasma in conscious human subjects: effects of hyperglycemia and hypoglycemia. *J Cereb Blood Flow Metab* 2002; 22: 271–279.
 31. Quistorff B, Secher NH and Van Lieshout JJ. Lactate fuels the human brain during exercise. *FASEB J* 2008; 22: 3443–3449.
 32. Van Hall G, Strømstad M, Rasmussen P, et al. Blood lactate is an important energy source for the human brain. *J Cereb Blood Flow Metab* 2009; 29: 1121–1129.
 33. Li H, Liu T, Peracchia C, et al. Properties and regulation of gap junctional hemichannels in the plasma membranes of cultured cells. *J Cell Biol* 1996; 134: 1019–1030.
 34. Berridge MJ. Neuronal calcium signaling. *Neuron* 1998; 21: 13–26.
 35. Stout C and Charles A. Modulation of intercellular calcium signaling in astrocytes by extracellular calcium and magnesium. *Glia* 2003; 43: 265–273.
 36. Zanotti S and Charles A. Extracellular calcium sensing by glial cells: low extracellular calcium induces intracellular calcium release and intercellular signaling. *J Neurochem* 2002; 69: 594–602.
 37. Rusakov DA. Depletion of extracellular Ca²⁺ prompts astroglia to moderate synaptic network activity. *Sci Signal* 2012; 5: pe4.
 38. Torres A, Wang F, Xu Q, et al. Extracellular Ca²⁺ acts as a mediator of communication from neurons to glia. *Sci Signal* 2012; 5: ra8.
 39. Xie L, Kang H, Xu Q, et al. Sleep drives metabolite clearance from the adult brain. *Science* 2013; 342: 373–377.